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Antioxidant vitamins increase the collagen content and reduce MMP-1 in a porcine model of atherosclerosis: implications for plaque stabilization

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Abstract

Degradation of extracellular matrix, particularly interstitial collagen, promotes plaque instability and contributes to restenosis after vascular injury. We have explored the effects of vitamins C and E on the collagen content and metalloproteinase-1 (MMP-1) expression after angioplasty in hypercholesterolemic pigs. Iliac angioplasty was performed on 18 minipigs divided into three diet groups: a normal-cholesterol (NC), a high-cholesterol (HC) and a high-cholesterol plus vitamins C+E (HCV). Four weeks later, after sacrifice, the vascular collagen content and MMP-1 protein expression, along with the plasma caseinolytic activity and lipid peroxidation, were measured. MMP-1 was also determined in arterial rings stimulated with native low-density lipoproteins (LDL) isolated from experimental groups. Cholesterol-rich diet augmented plasma lipid peroxidation (P < 0.05), reduced the collagen content and increased vascular MMP-1 expression after injury (P < 0.05). Enhanced caseinolytic activity (identified as MMP-1) was also observed in HC plasma samples and in supernatants from arterial rings incubated with HC-LDL. Vitamins C and E markedly increased neointimal collagen content (P < 0.01), reduced the hypercholesterolemia-induced changes in vascular MMP-1 (P < 0.05) and diminished plasma and ex vivo caseinolytic activity. Vitamins C and E may help stabilize atherosclerotic plaque after angioplasty and favor vascular remodeling by increasing collagen content and reducing vascular MMP-1 expression in porcine hypercholesterolemia.

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1. Introduction

Degradation of extracellular matrix (ECM) contributes crucially to a variety of processes, including atherosclerosis and restenosis after vascular injury and is a critical event leading to the weakening of the vessel wall [1,2]. ECM degradation by metalloproteinases (MMPs) is known to be involved in the pathogenesis of cardiovascular diseases, including atherosclerosis, restenosis and myocardial infarction [3]. MMPs are a family of zinc-dependent enzymes that can be divided

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into two structurally distinct groups, such as secreted MMPs and membrane-type MMPs. Secreted MMPs include collagenases (MMP-1, MMP-8), gelatinases (MMP-2, MMP-9) stromelysins (MMP-3, MMP-10) and other MMPs. MMP-1 secreted from cells present in atheroma could be involved in the subendothelial destruction of ECM and remodeling of plaques [3,4].

Previous studies have demonstrated that hypercholesterolemia induces local collagen loss, contributing to plaque destabilization [5]. Although the mechanisms involved in these processes are still unclear, it has been suggested that the activation of cells in atheroma could be a principle source of MMPs. oxLDL stimulate macrophage activation and induce the expression of MMP-1 by vascular cells [6]; this proteinase is known to play a prominent role in plaque stability [7] and, in

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addition, lipid lowering has been shown to reduce MMP-1 activity and increase collagen content of atheroma [8].

Antioxidant vitamins have been evaluated through clinical studies for their association with atherosclerotic disease, but their results are not conclusive. Recent clinical trials enrolling patients with established atherosclerosis or at high vascular risk, have failed to demonstrate that vitamin E reduces the progression of the disease [9]. In contrast, other studies have demonstrated that antioxidant vitamins retard the development of coronary arteriosclerosis [10]. In experimental models, we and others have previously shown that antioxidants are able to reduce intimal lesions, decrease type-1 plasminogen activator inhibitor (PAI-1) after injury and restore endothelial function [11–13], but their possible effect on the regulation of MMPs and/or neointimal collagen content has not been properly assessed.

The pig model appears to be the most relevant to human restenosis because of its similarities with human coronary circulation, spontaneous development of atherosclerosis and histological response to vascular injury [14]. We hypothesized that antioxidant vitamins C and E would preserve the collagen content in a hypercholesterolemic porcine model of angioplasty, possibly by reducing the MMP-1 expression. Treatment with vitamins was initiated 3 weeks after the induction of hypercholesterolemia in order to more closely mimic a potential clinical situation.

2. Material and methods

2.1. Animals

Eighteen Yucatan miniature pigs (4 months old, mean weight 33.29 ± 1.69 kg) were obtained from our breeding center and maintained in the animal facilities at CIFA (Centro de Investigación en Fármaco-Biología Aplicada), a GLP-accredited center within the University of Navarra (Pamplona, Spain). Housing conditions followed the European regulations (86/609/CEE).

2.2. Biochemical measurements

Concentrations of total serum cholesterol and its fractions were determined by standard enzymatic assays.

Plasma lipid peroxidation was measured as thiobarbituric acid reactive substances (TBARS) [15]. Vitamin E was measured in plasma using a reverse-phase high-performance liquid chromatography (HPLC) [13].

2.3. Vascular injury and tissue processing

Our model of vascular lesion after iliac angioplasty in hypercholesterolemic pigs has recently been described [11]. Briefly, animals were divided into three groups (n = 6) and fed different dietary formulations for 4 weeks: a normal-cholesterol (NC) control group, given standard porcine chow (Porcisanders, Sanders); a highcholesterol (HC) group fed with a diet containing 24.5% animal lard, 4% cholesterol (Roig Farma) and 1.5% biliar extract (Roig Farma); and a high-cholesterol plus vitamins C and E (HCV) group fed similar to the HC group, but supplemented during the last week with 1 g vitamin C and 1000 IU vitamin E (Roig Farma) per animal per day. The dosage of antioxidants was chosen from a similar model demonstrating that animals treated with vitamins C and E have a smaller degree of LDL oxidation and the best response to vascular injury [12]. At this time, the animals were subjected to catheterization in the right internal iliac artery, the left internal iliac artery was used as control. The animals from each group were allowed to recover and maintained their designated diets until they were sacrificed, 28 days after interven-

Vascular tissues were fixed with paraformaldehyde 4%, dehydrated and paraffin-embedded for histological purposes. Arterial blood samples were drawn before and after arterial injury for analytical measurements.

2.4. Immunohistological analysis

Sirius red staining was used to identify interstitial collagen on serial 3 μ m sections as previously described [16].

Tissue sections for MMP-1 immunohistochemistry were heated in sodium citrate buffer (0.01 M, pH = 6) for antigen retrieval before their incubation with 2 μ g/ml murine anti-MMP-1, which recognizes both active and latent forms (IM35L Oncogen, USA), 0.5 μ g/ml murine anti- α -smooth muscle actin (M850, Dako, Denmark) and murine anti-swine leukocyte Ag monoclonal antibody 4E9 [17] (kindly provided by Dr J. Dominguez, INIA, Spain). All sections were incubated with secondary biotinylated rabbit anti-mouse Fc (Dako), developed with avidin-biotin-peroxidase complex (ABC, Dako, Denmark) and 3-3′-diaminobenzidine hydrochloride (Sigma, USA) and counterstained with Harris' hematoxylin [11].

A visual grading scale was used for assessing the intensity of MMP-1 immunostaining. Three observers, independently and in a blinded fashion, characterized sections by comparing them against a control section using the grading scale. Each observer evaluated and scored each field on a 0 to 3+ scale for the extent distribution of positive staining [11].

2.5. Effect of porcine LDL on MMP-1 ex vivo expression

Coronary artery rings (n = 9) were incubated for 48 h with native LDL (100 µg protein/ml) isolated from

bisochemical measurements and morphometric analysis of iliac arteries in the porcine model of angioplasty 28 days after injury

Group	Total-cholesterol (mM)	Vitamin E (µM)	Vitamin E/total-cholesterol	TBARS (nM)	Intimal area (mm ²)	Stenosis (%)	Stenosis (%) Media cell density (10 ² cells/mm ²)
•	,						
NC	1.6 ± 0.1	1.9 ± 0.9	1.11 ± 0.41	357.9 ± 49.4	1.2 ± 0.5	42.2 ± 12.9	38.6 ± 1.5
HC	$5.9 \pm 1.5**$	1.8 ± 0.5	0.30 ± 0.03	$553.4\pm66.9**$	1.7 ± 0.6	26.6 ± 4.6	49.6 ± 2.3
HCV	$5.4 \pm 5.4**$	$15.9\pm1.9**,^{\dagger\dagger}$	$3.03 \pm 0.32^{*,\dagger\dagger}$	$212.5\pm43.7*$	$0.8\pm 0.1^{\dagger}$	28.6 ± 2.7	35.4 ± 2.7

* Differences from NC as P < 0.05

** Differences from NC as P < 0.01† Differences from HC as P < 0.05.

 †† Differences from HC as P < 0

porcine plasma from experimental groups (NC, HC and HCV) by density gradient ultracentrifugation [18]. Supernatants of cultured rings were separated by SDS-PAGE under reducing conditions in Tris-glycine-SDS buffer (pH 8.3), electroblotted onto nitrocellulose membrane and probed with murine anti-MMP-1 (1 μ g/ml), followed by a peroxidase-linked rabbit anti-mouse IgG (1:2000, Zymed, USA). Blots were developed using a chemiluminescence system (ECL, Amersham-Bioscience), on X-ray film. Human recombinant MMP-1 (Amersham Bioscience) was used as the positive control.

2.6. Zymographic analysis

Some 10 µl of coronary supernatants or a 1:20 dilution of plasma samples in saline were loaded on a standard 8% SDS-PAGE gel containing 0.5% casein and electrophoresed under non-reducing conditions. Substrate gels were soaked with Triton X-100 solution (2.5%) for 15 min at room temperature and then incubated overnight at 37 °C with a 50 mM Tris-HCl buffer (pH 7.4, 0.05% NaCl, 0.1% NaN₃ and 10 mM CaCl₂). The lysis of the casein in the gel was visualized by staining with Coomassie brilliant blue (Sigma), densitometrically quantified on a GS690 system (Bio-Rad) and identified on the basis of protein electrophoretic mobility [19]. Quenching analysis were performed by simultaneous incubation of plasma samples with 5-50 ng of a monoclonal antibody anti-MMP-1 (Oncogene).

2.7. Statistical analysis

All data was expressed as mean \pm S.E.M. Statistically significant differences between groups were calculated by analysis of variance (ANOVA) followed by Tukey posthoc test or Kruskal–Wallis and Mann–Whitney test, for parametric and non-parametric conditions, respectively. Statistical significance was accepted at the 95% confidence level (P < 0.05). Association between variables was assessed by using the Spearman's correlation test. Analysis was performed by using the computer program SPSS for Windows (SPSS Inc).

3. Results

3.1. Plasma cholesterol, lipid peroxidation and lesion formation

As shown in Table 1, pigs receiving hypercholesterolemic diet showed higher cholesterol concentration (P < 0.01) and plasma lipid peroxidation (P < 0.01)than the controls. The angioplasty induced severe structural changes in the vascular wall in comparison

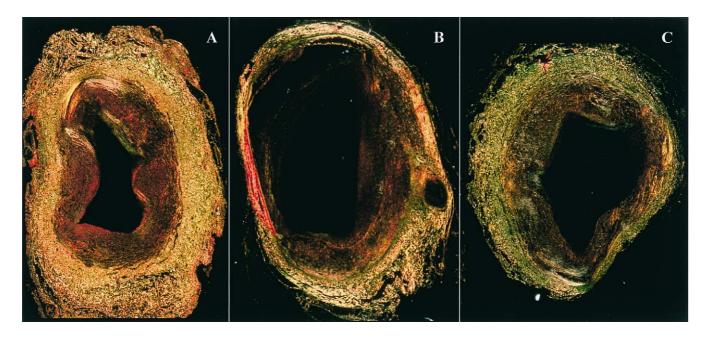


Fig. 1. Sirius red staining of vascular tissue sections from the three experimental groups, photographed with a polarizing filter. Intense collagen loss was observed in the injured HC arteries (panel B) in comparison with the NC group (panel A). Treatment with vitamins C and E induced a significant increase in the collagen content of vascular sections (panel C).

with uninjured arteries, resulting in an increased intimal area and media cell density in HC group compared to the NC group (data not shown).

Treatment with vitamins C and E had no effect on total cholesterol or LDL concentration, but, as expected, it increased plasma vitamin E levels and, particularly, the vitamin E content in the LDL fraction compared with HC (14.5 ± 3.6 vs. 4.29 ± 2.1 nM/mg protein, P < 0.05). As shown in Table 1, antioxidant treatment significantly reduced the hypercholesterolemia-induced plasma lipid peroxidation at the time of sacrifice (P < 0.01), without significant improvement in morphologic parameters, except a slight, but significant, reduction in the intimal area (P < 0.05).

3.2. Atherosclerotic lesion collagen content

Vascular sections stained for collagen with Sirius red were analyzed under polarized light. As shown in Fig. 1, the lesions in the NC group were intensely stained for collagen fibers forming intricate networks throughout the plaque (Fig. 1A). However, the collagen content was practically absent throughout the vascular wall in the HC group (Fig. 1B), whereas a significant recovery was observed in the HCV group (Fig. 1C). The collagen stained area in plaques was 2.4-fold higher in the NC group than in HC group ($70.8\pm8.1\%$ vs. $29.3\pm8.9\%$, P < 0.01). Antioxidant vitamins C and E significantly

restored the collagen content with respect to HC (65.5 \pm 7.4% vs. 29.3 \pm 8.9%, P < 0.01) without differences regarding the NC group.

3.3. MMP activity

Since no vascular samples were available for zymography, we performed caseinolytic analysis of plasma samples in order to determine whether hypercholesterolemia induces increased systemic proteolytic activity. A similar proteolytic profile activity was observed in all of the experimental groups, but a more intense 52 kDa proteolytic band was found in the samples obtained after injury in the HC group than in the NC group (P <0.05, Fig. 2A). However, as shown in Fig. 2(A), the 52 kDa caseinolytic activity was reduced in the group treated with antioxidants in comparison to the HC group, indicating that in addition to local effects on collagen content, vitamins C and E also reduced the systemic proteolytic activity. In order to identify the caseinolytic band, additional experiments were performed by preincubating plasma samples with different concentrations of specific MMP-1 antibody. As shown in Fig. 2(B), a dose-dependent reduction in the proteolytic activity was found, indicating that it corresponds to MMP-1.

Taking a further step, we performed immunohistochemical analysis for MMP-1 in vascular sections in

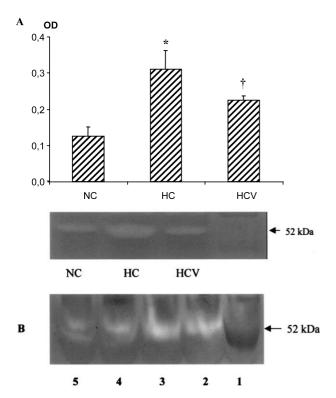


Fig. 2. Caseinolytic activity of plasma samples and quantitative analysis (top) of proteolytic activity in NC, HC and HCV groups (A). Quenching experiments performed by simultaneous incubation of plasma samples with different concentrations (5–50 ng) of a monoclonal antibody anti-MMP-1 show the specificity of the caseinolytic activity (B). Lane 1: molecular weight marker; lane 2: HC plasma sample; lane 3–5: plasma sample preincubated with 5, 25 and 50 ng of murine anti-MMP-1 antibody respectively. *P < 0.05 in comparison with HC group.

order to assess whether MMP-1 expression could be associated with observed modifications in collagen content within vascular vessels in the experimental groups. As shown in Fig. 3, MMP-1 protein was significantly increased in the neointima and media layers of HC group compared to NC (P < 0.05, Fig. 3A, B). Median quantitative score for MMP-1 immunostaining was also found to be significantly higher (P < 0.05) in the HC group (Table 2). The most intense signal was observed within the vascular lesion, near the media and in the thickened intima (Fig. 3D). Areas with the most intense signal were also positive for α -actin smooth muscle cells (SMC) and macrophages, suggesting that these cellular types are involved in MMP-1 vascular expression (Fig. 3E, F).

Interestingly, the antioxidant treatment was able to significantly reduce the observed vascular MMP-1 expression in response to injury in the HCV group. Immunohistochemical analysis showed weaker MMP-1 reactivity in the media and in the thickened intima in the group of pigs receiving antioxidants than in the HC group (P < 0.05) (Fig. 3C). Median quantitative score

for MMP-1 immunostaining was found to be significantly lower (P < 0.05) in the HCV group than in the HC group (Table 2).

3.4. Effect of native LDL on MMP-1 activity and expression

Ex vivo experiments performed with supernatants from cultured porcine coronary rings showed an increased 52 kDa caseinolytic activity in the segments incubated with LDL obtained from the HC group (Fig. 4A). A weaker ex vivo proteolytic activity could be observed in the conditioned medium from coronary rings treated with HCV-LDL than those treated with HC-LDL; the said activity was later identified as MMP-1 by Western blot analysis (Fig. 4B).

4. Discussion

Degradation of ECM is a critical event leading to the weakening of the vessel wall. A mechanistic link between hypercholesterolemia and collagen loss is still hypothetical, although a critical role of MMPs and other proteolytic enzymes have also been strongly suggested [5]. In the present study, we report that a combined dietary treatment of vitamins C and E restores the hypercholesterolemia-induced collagen loss and reduces both vascular and systemic MMP-1 after vascular injury in pigs. This effect was largely independent of lipid lowering and occurred despite trivial morphologic changes in the vessel wall.

We had previously shown that hypercholesterolemia induces severe structural and morphologic changes in the vessel wall after angioplasty [11]. This study extends those findings by showing that Sirius red stained injured arteries from hypercholesterolemic pigs exhibit extensive areas devoid of collagen filaments (coinciding with recent reports [20]) suggesting an increased proteolytic activity or diminished collagen synthesis. The zymographic analysis of plasma samples displaying an intense MMP-1 caseinolytic band as well as an increase in vascular MMP-1 protein expression would support those findings. Areas positively stained for MMP-1 were observed mainly in the media and in the intimal thickening close to internal elastic lamina in the HC group, thereby confirming previous reports [21–23]. MMP-1 has been shown to be specifically present in atherosclerotic lesions [6,24], secreted by endothelial cells, SMC and macrophages and could be involved in the ECM degradation, which is considered an important feature for plaque stability [7,25]. Our results show that areas with the most intense signal were also positive for SMC and macrophages, confirming that both cellular types are involved in the MMP-1 vascular expression.

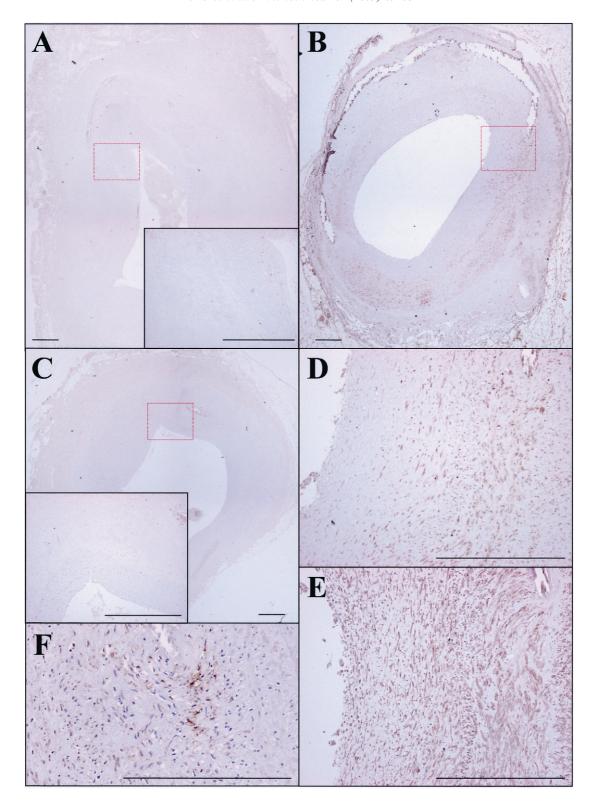


Fig. 3. Immunohistochemical pattern for vascular MMP-1 in the different groups of pigs. Whereas a weak MMP-1 signal (score = 1) was present in NC (panel A) in comparison with the HC (score = 3) (B), the vascular immunoreactive MMP-1 expression after angioplasty was significantly reduced (score = 1) in hypercholesterolemic pigs treated with vitamins C and E (panel C), thereby being similar to NC. Areas squared in red correspond to magnified details. Panel D shows a magnified detail of (B). Panels E and F show the smooth muscle α -actin (E) and macrophages (F) staining of the same region. Scale bar, 400 μ m.

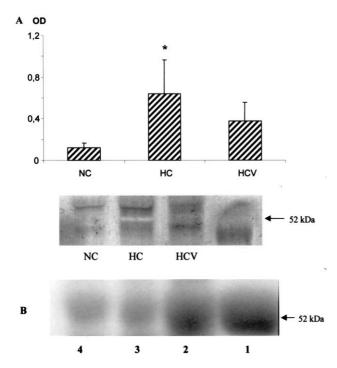


Fig. 4. Effect of purified LDL on MMP-1 induced in coronary artery rings. Caseinolytic activity of supernatants from LDL-stimulated porcine rings and quantitative analysis (top) of caseinolytic activity in NC, HC and HCV groups (A). Western blotting for MMP-1 in supernatants of cultured aortic rings (B). Lane 1: standard of human recombinant MMP-1 (52 kDa). Supernatants from cultures incubated with LDL from HC (lane 2), HCV (lane 3) and NC (lane 4), all three showing a positive band at 52 kDa size.

Table 2 Semiquantitative score assessment of vascular MMP-1 in the porcine iliac artery by immunohistochemistry

Group	Group size	Score				Median score
		0	1	2	3	
NC	6	2	3	1	0	1
HC	6	0	0	3	3	2.5*
HCV	6	0	4	2	0	1

^{*} P < 0.05 in comparison with both NC and HCV.

In a further step, we performed ex vivo experiments in cultured normal coronary rings to assess the effect of native LDL, from the three experimental groups, on MMP-1. The fact that MMP-1 was increased after stimulation with LDL from hypercholesterolemic pigs would support the idea that hypercholesterolemia plays a direct role in the induction of MMP-1 by vascular cells. According to this data, hypercholesterolemia would play a significant role in the secretion of MMP-1 by cells present within the atheroma.

For the first time, we provide evidence that antioxidant vitamins C and E, despite a minimum effect on morphologic changes in the vascular wall, can signifi-

cantly increase the collagen content in hypercholesteromic injured arteries, which could be related to a significant reduction of vascular MMP-1. That effect was independent of lipid lowering because vitamin treatment did not alter plasma cholesterol levels. Moreover, the systemic proteolytic activity was also attenuated in the plasma of the pigs receiving antioxidants. Finally, ex vivo incubation of porcine coronary arteries with native LDL from hypercholesterolemic pigs treated with vitamins C and E attenuated the enhancement of caseinolytic activity induced by hypercholesterolemia. Based on aforementioned data together, it appears that the combination of vitamins C and E could reduce the hypercholesterolemia-induced MMP-1 expression in our experimental model of vascular injury and, therefore, prevent ECM degradation by increasing the collagen content, thus contributing to plaque stabilization. Other studies have reported a slow progression of atherosclerosis in experimental models using antioxidant vitamins C and E [11,12,26]. In addition, it has been recently shown that vitamin C deficiency induces a type of plaque morphology that is potentially vulnerable to rupture [27].

The mechanism by which antioxidants may participate in matrix-protective processes remains unclear. We show herein that the combination of vitamins C+E reduces plasma lipid peroxidation. Oxidative stress is increasingly being recognized as a potentially important contributor to atherogenesis and restenosis after vascular interventions and injury [28,29]. The generation of reactive oxygen species and oxidation of lipids have profound and wide-range effects that can dramatically increase vascular toxicity and initiate a cascade of molecular and cellular responses. Increased vascular expression of MMP-1 in hypercholesterolemia can be a direct effect of oxidative stress because reactive oxygen species are known to enhance MMP-1 expression and activity in vitro [30,31]. Our results, which show that dietary supplementation with vitamins C+E was able to significantly reduce plasma lipid peroxidation, support this hypothesis. In fact, constituents of oxidatively modified lipoproteins that stimulate protein kinase C (PKC) might link to the activator protein-1 (AP-1)mediated transcriptional regulation that plays a key role in controlling transcription of MMP-1 [8]. Moreover, NF-κB, a transcription factor activated by oxidative stress/free radicals [32], has been shown to be required for cytokine upregulation of MMP-1 in vascular cells [33]. α -Tocopherol is a PKC inhibitor capable of diminishing MMP-1 expression in fibroblast cultures [34], but it can also inhibit NFkB, resulting in a significant inhibition of monocyte cell adhesion or cytokine regulation, which is required for upregulation of MMP-1 in in vitro studies [33,35].

Our study has some limitations. Other MMPs are also playing a role in arterial remodeling after injury and ECM degradation. In immunohistochemical staining, the presence of MMPs assessed by immunohistochemistry does not imply increased catalytic capacity [23]. In addition, matrix catabolism in the cardiovascular system involves enzymes other than the MMP family, such as cathepsins and cystatins.

In conclusion, the present study is the first to show an increase in the neointimal collagen content and a reduction of vessel-wall MMP-1 in a porcine hypercholesterolemic model of vascular injury in response to vitamins C and E, thereby suggesting that antioxidant therapy can contribute to prevent matrix degradation in atherosclerosis, thus contributing to plaque stabilization.

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